Case-control studies in epidemiological research

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- In a case-control study design two groups of subjects, one with the disease (or a health outcome) and the other without the disease are included. Exposure information are collected from these subjects. The subjects with the disease are referred to as cases while the subjects without the disease are referred to as controls.
- The cases and controls are not selected randomly from the underlying population. Rather, case subjects are, generally, randomly selected from the population of diseased people and controls are randomly selected from the population of disease-free people. Importantly, the sampling must not depend on subjects' exposure status.
- The purpose of the case-control study is to find the connection/association/effect of exposure on the disease.

- The design is highly efficient, it can be done in a much shorter time period (less costly) compared to a similar cohort study.
- The design can be used to study rare and common diseases.
- If an exposure is rare in the population, then whether we use a case-control or cohort study, both designs need a large sample size. However, if the exposure is strongly associated with the disease, then a case-control study will be much more efficient than a cohort study.
- In a case-control design, cases and controls must be comparable, otherwise results are biased.
- Since the exposure information is collected retrospectively, the measurements are subject to recall bias that may lead to bias results.

- Suppose that we have a case-control data with a scalar dichotomous exposure variable, call it X. Let Y be the disease variable taking on zero and one for control and case subjects, respectively.
- The data can then be summarized as follows:

	Unexposed Exposed		
	(X = 0)	(X = 1)	
Control	а	Ь	n 0
Case	с	d	n_1

- n_0 : total number of controls
- n_1 : total number of cases

Two common measures of associations: relative risk (RR) and odds ratio (OR)

• RR = pr(Y = 1|X = 1)/pr(Y = 1|X = 0), a ratio of the risk of the disease from exposed to unexposed, generally RR is not estimable from case-control data

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OR =
$$\frac{\text{Odds of the disease among exposed}}{\text{Odds of the disease among unexposed}}$$

= $\frac{\text{Odds of exposure among disease}}{\text{Odds of the exposure among non-disease}}$

• Fortunately, OR is estimable from a case-control data, and its estimator is

$$\widehat{OR} = \frac{ad}{bc}.$$

- When the exposure and disease are independent, RR = 1 and OR = 1.
- In the presence of dependence, $RR \neq 1$ and $OR \neq 1$.
- Thus, testing of independence is equivalent to test OR = 1 or log(OR) = 0.
- *OR* > 1 : Odds of the disease is greater in the exposed group than in the unexposed group, exposure is a risk factor
- *OR* < 1 : Odds of the disease is less in the exposed group than in the unexposed group, exposure is a protective factor

A dataset on average daily consumption of alcohol and oesophageal cancer $^{\rm 1}$

	0-79 g	80+ g	
	(X = 0)	(X = 1)	
Control	666	109	775
Case	104	96	200

•
$$\widehat{OR} = 666 \times 96/104 \times 109 = 5.64$$

- The standard error of $\widehat{\log}$ is $\sqrt{666^{-1} + 109^{-1} + 104^{-1} + 96^{-1}} = 0.17$.
- The 95% large sample CI of the OR is exp{log(5.64) ± 1.96 × 0.17} = (3.99, 7.92).

 ¹ Statistical Methods in Cancer Research by Breslow and Day, (1980, ≡p. 123. ≡
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Code

```
# Uploading the necessary package
library(epitools)
# Entering the data
  data=array(c(666, 104, 109, 96), dim=c(2, 2),
  dimnames=list(y=c(0, 1), x=c(0, 1)))
> data
    x
y 0 1
0 666 109
1 104 96
```

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```
> oddsratio(data)
$data
      х
          0
             1 Total
у
       666 109
                  775
 0
  1
       104 96
               200
 Total 770 205
               975
$measure
   odds ratio with 95% C.I.
   estimate lower
y
                      upper
 0 1.000000
                  NA
                            NA
  1 5.624773 3.992007 7.947159
$p.value
  two-sided
   midp.exact fisher.exact
                           chi.square
y
 0
           NA
                         NA
                                      NA
  1
            0 1.079486e-22 8.614569e-26
$correction
[1] FALSE
```

- The odds of the disease (or cancer) among the exposed group is 5.64 times that of among the unexposed group.
- If we know, the disease is rare (less than 1% in the population), then the odds ratio can be interpreted as the relative risk. Then we can say, the risk of the disease among exposed group is 5.64 times that among the unexposed group.
- We <u>do not claim</u> that (3.99, 7.92) includes the true OR with 0.95 probability. Rather, in repeated sampling from the underlying population and computation of the 95% CI, about 95% of the intervals include the true OR. However, we do not know if (3.99, 7.92) includes the true OR nor we can associate any probability with this interval. Just say that we are 95% confident that the true odds ratio is in (3.99, 7.92).

- The above example indicates the risk of the esophageal cancer among exposed group is 5.64 times that among the unexposed group.
- Let's look at this more closely. Let pr(Y = 1|X = 1) = 0.00056 and pr(Y = 1|X = 0) = 0.0001, and the relative risk in this case 5.6. In another situation, pr(Y = 1|X = 1) = 0.28 and pr(Y = 1|X = 0) = 0.05, this also results in the relative risk of 5.6. In one case, the expected number of cancer cases increases from from 1 to 5.6 when 10,000 unexposed cases all become exposed. In the second case, the expected number of increase is from 5 to 28 when 100 unexposed subjects all become exposed.
- Obviously, assuming that the risk is a causal, the second case is more of a public health concern than the first case.
- However, only the relative risk is not able to shade light on this entire situationparticularly, it does not tell us the gravity of the problem in the view of public health.

Population attributable risk (PAR) measures **how much of the disease in the population is caused by the risk factor**. PAR is measured as the excess rate of disease in the total study population of exposed and unexposed individuals compared to the population of unexposed individuals, and is formulated as

 $PAR = \frac{Risk \text{ of the disease in total population} - Risk \text{ of the disease among the unexposed}}{Risk of the disease in total population}$

In terms of notations,

$$PAR = \frac{\operatorname{pr}(Y=1) - \operatorname{pr}(Y=1|X=0)}{\operatorname{pr}(Y=1)}$$

It can also be expressed as

$$\mathrm{PAR} = \frac{P_e(R-1)}{P_e(R-1)+1},$$

where P_e stands for the proportion of population exposed and R stands for the relative risk of the disease from exposed to unexposed.

- For the esophageal cancer, we may assume that 1) the control population is very similar to the general population, and 2) controls are sampled randomly from the populations. Then P_e can be estimated by the proportion of exposed in the control sample, 0.14. Thus, $\widehat{PAR} = 0.14(5.6 1)/\{0.14(5.6 1) + 1\} = 0.39$.
- Thus, about 39% of the disease in the general population is caused by the exposure (average daily alcohol consumption 80+ g).

- Although simple, the analysis of a 2 × 2 table is a over simplification, and may risk a confounding bias. For our toy example, the age effect likely be confounded in the disease-exposure association.
- Suppose now that the data are stratified for different age groups. It is presented in the next slide.

Several 2×2 tables

		0-79 g	80+ g	
Age		(X = 0)	(X = 1)	$\widehat{\mathrm{OR}}$
25-34	Control	106	9	∞
	Case	0	1	
35-44	Control	164	26	5.05
	Case	5	4	
45-54	Control	138	29	5.67
	Case	21	25	
55-64	Control	139	27	6.36
	Case	34	42	
65-74	Control	88	18	2.58
	Case	36	19	
75+	Control	31	0	∞
	Case	8	5	

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- It is clear that the estimated conditional odds ratios are changing. Are the underlying true conditional odds ratios different?
- To check this, test the homogeneity of odds ratios (Breslow-Day test). Let θ_k be the conditional odds ratio for age group k.
- Set H_0 : $\theta_1 = \cdots = \theta_6$ versus H_a : at least one of θ_k 's is different from the rest.

Code

```
# Uploading the necessary package
library(DescTools)
# Data input
data2=array(c(106, 0, 9, 1, 164, 5, 26, 4, 138, 21, 29, 25, 139, 34, 27,
42, 88, 36, 18, 19, 31, 8, 0, 5), dim=c(2, 2, 6), dimnames=list(y=c(0, 1)
x=c(0, 1), age=c("25-34", "35-44", "45-54", "55-64", "65-74", "75+")))
BreslowDayTest(data2)
```

Breslow-Day test on Homogeneity of Odds Ratios

```
data: data2
X-squared = 9.3234, df = 5, p-value = 0.09684
```

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- Since the *p*-value is large, we fail to reject *H*₀ at the 5% level, and conclude that there is not enough evidence that the underlying conditional odds ratios are different (seems contradictory, isn't it?).
- Important point is that if cell frequencies are small, then Breslow-Day (BD) test is not very reliable. Then one should fit a logistic regression model and then carry out the test.
- Since BD test was non-significant at the 5% level, we now aim to estimate that common conditional odds ratio. This odds ratio is adjusted for age.

Mantel-Haenszel estimate:

$$\widehat{\theta}_{mh} = \frac{\sum_{i} a_{i} d_{i} / N_{i}}{\sum_{i} b_{i} c_{i} / N_{i}}$$

- +: It is not affected by zero cell frequencies
- -: The standard calculation is very tedious
- Next slide we show how to obtain the MH estimate and conduct the hypothesis test of conditional independence of the disease and the exposure.

```
# Uploading the necessary package
library(stats)
```

```
mantelhaen.test(data2)
```

```
Mantel-Haenszel chi-squared test with continuity correction
```

- The age adjusted conditional odds ratio is 5.2 with a 95% CI (3.6, 7.5).
- With the *p*-value smaller than 0.05, we reject *H*₀, and conclude that there is a strong conditional association between the disease and heavy alcohol consumption.

- When the conditional odds ratios are the same across the strata, and they are equal to the crude odds ratio, the potential confounding variables do not have any confounding effect.
- When the conditional odds ratios are the same across the strata, and they are not equal to the crude odds ratio, the potential confounding variables carry confounding effect.
 - If the common conditional odds ratio is larger than the crude odds ratio, the confounder is termed as negative confounder.
 - If the common conditional odds ratio is smaller than the crude odds ratio, the confounder is termed as positive confounder.
- When the conditional odds ratios are not the same across the strata, we call the confounder as an effect modifier.

²M.C. Costanza, Matching, Preventive Medicine. 1995; 24:425–433

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Why?

- For very small cell frequencies, BD test for testing homogeneity of odds ratio is not reliable.
- The number of confounding variables may not be a scalar or categorical variable. In the presence of a large number of confounding variables, MH method or BD test is not practical.

These issues can be overcome by fitting a logistic model. Of course, it requires some parametric model assumptions.

- High level of daily alcohol consumption: X = 1, not high level of alcohol consumption: X = 0
- Age: confounding variable (Z), treat it as a numeric with Z = 1 for age 25-34, Z = 2 for age 35-44 etc.
- Model π , the probability of Y = 1 given X and Z:

$$logit(\pi) = \alpha_0 + \alpha_1 X + \alpha_2 Z + \alpha_3 X Z$$

• In the above model the age effect is assumed to be linear (a parametric assumption).

- If the interaction term α₃ is zero, then the conditional odds ratios are homogeneous. So, test of homogeneity can be done by testing H₀ : α₃ = 0 versus H_a : α₃ ≠ 0.
- If the above test is statistically significant, then it is meaningless to estimate the age adjusted common odds ratio (because there is no such common odds ratio). Rather, we should report the conditional odds ratio for each age group. Also, the confounding variable is termed as an effect modifier.

- If the above test is statistically non-significant, then we can report the age adjusted common odds ratio, exp(α₁).
- We can make statistical inference on this conditional odds ratio.
- The logistic model can include many confounders.

```
# Preparation of the data in the desirable format
mydata=as.data.frame.table(data2)
disease= mydata$Freq[seq(2, 24, 2)]
total=mydata$Freq[seq(1, 24, 2)]+ mydata$Freq[seq(2, 24, 2)]
exposure= mydata$x[seq(2, 24, 2)]
age= as.numeric(mydata$age[seq(2, 24, 2)])
```

```
# Invoking the GLM function to fit the logistic model
out=glm(disease/total~exposure+age+exposure*age, family=binomial,
weight=total)
summary(out)
```

Deviance Residuals:					
Min	1Q	Median	ЗQ	Max	
-2.58762 -1.	72026	0.08124	1.19703	1.49961	
Coefficients:					
	Estimat	e Std. Er	ror z valu	1e Pr(> z)	
(Intercept)	-4.0912	0.36	5114 -11.32	29 < 2e-16	***
exposure1	1.7510	0.63	839 2.74	13 0.00609	**
age	0.6136	0.08	531 7.19	93 6.32e-13	***
exposure1:age	0.0077	9 0.16	424 0.04	17 0.96217	
(Dispersion parameter for binomial family taken to be 1)					
Null deviance: 211.608 on 11 degrees of freedom					
Residual deviance: 31.929 on 8 degrees of freedom					
AIC: 80.257					

- The interaction term is statistically non-significant at the 5% level.
- The age adjusted common odds ratio estimate of the disease-exposure association is exp(1.75) = 5.75 with a 95% Cl $exp(1.75 \pm 1.96 \times 0.64) = (1.64, 20.17)$.
- Why does this OR estimate differ from the MH estimate?
- For a case-control data, the intercept term has no useful interpretation.
- For any given exposure group, the odds ratio of the disease for 10 years increase in age is exp(0.61) = 1.84. In other words, for any given exposure group, the odds of the disease increases by 84% for 10 years increase in age.

- As a strategy to control confounding, often matched data are analyzed. Matching can be introduced at the design stage or at the data analysis stage. In essence, for every case, a number of controls are chosen/included in the study by matching the values of a set of potential confounding variables.
- If the matching is introduced at the design stage, then we can keep a constant ratio of cases to controls in each stratum defined by the matching variables. If the matching is done before the analysis (not at the data collection stage), then there is a possibility of imbalance in the ratio of cases to controls across the strata.
- Two basic objectives of matching: better control of the confounding effect and increase the efficiency of measuring the strength of the association
- For the analysis of matched case-control data, conditional logistic regression method is used.

A toy example of matched data 3

Code

library(epiDisplay)						
	data(VC1to6)					
>	> head(VC1to6)					
	matset	case	smoking	rubber	alcohol	
1	. 1	1	1	0	0	
2	: 1	0	1	0	0	
З	2	1	1	0	1	
4	. 2	0	1	1	0	
5	3	1	1	1	0	
6	3	0	1	1	0	

Matching variables: age, sex, neighborhood

rubber: worked in the rubber industry

³Chongsuvivatwong, V. 1990 A case-control study of esophageal cancer in Southern Thailand. J Gastro Hep 5:391–394.

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R code for the conditional logistic analysis

```
library(survival)
out5=clogit(case<sup>-</sup>smoking+rubber+alcohol+strata(matset), data=VC1to6)
> summary(out5)
Call:
coxph(formula = Surv(rep(1, 119L), case) ~ smoking + rubber +
   alcohol + strata(matset), data = VC1to6, method = "exact")
 n= 119, number of events= 26
       exp(coef) exp(-coef) lower .95 upper .95
smoking 1.5523 0.6442 0.4375 5.508
rubber 0.6331 1.5797 0.1780 2.251
alcohol 5.2951 0.1889 1.6490 17.003
Concordance = 0.688 (se = 0.065)
Likelihood ratio test= 12 on 3 df, p=0.007
Wald test = 9.18 on 3 df, p=0.03
Score (logrank) test = 11.24 on 3 df, p=0.01
```

- After adjusting the effect for the confounding variables (age, sex, neighborhood), and other risk factors, smoking and rubber, the odds ratio of association between cancer and alcohol is 5.3.
- Note that smoking and rubber are not confounding variable, they are just potential co-risk factors. The data were also analyzed by including alcohol × smoking and alcohol × rubber interaction terms, and both terms turned out statistically non-significant. This indicates that they are not effect modifiers.

- Researchers are worried that matching and conditional analysis does not reduce the confounding bias always. Sometimes, matching may introduce bias.⁴
- If all potential confounding variables are measured, then matched data can be used to estimate the causal odds ratio of association.⁵
- Observational studies need not be cohort or case-control studies. There are many variations, and among them nested case-control study (case-controls are nested within a cohort) is popular choice. It is efficient, and provides more information on the disease etiology, such as the incidence rate of the disease with the chronological age of the subjects and its association with the risk factors, compared to a simple case-control study.⁶

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⁴M.C. Costanza, Matching, Preventive Medicine. 1995; 24:425–433.

 $^{^{5}}$ Rose S, Laan MJ. Why match? Investigating matched case-control study designs with causal effect estimation. *Int J Biostat.* 2009;5(1):1

⁶V.L. Ernster, Nested Case-Control Studies, *Preventive Medicine*. 1994; 23:587–590، م

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Thank you!

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